

TIU Transactions on Human sciences

# Unveiling Nature's Detox: Microbial Arsenic Bioremediationfor Human Health: A Review

Arindam Chakraborty<sup>1†</sup>, Arnab Ganguli<sup>1</sup>

<sup>1</sup> Techno India University, West Bengal, 700091, India

#### Abstract

The review paper, "Unveiling Nature's Detox: Microbial Arsenic Bioremediation for Human Health," thoroughly examines the global health threat posed by arsenic contamination, emphasizing its chronic and acute dangers. The paper explores microbial arsenic bioremediation mechanisms, revealing the ingenious strategies employed by microorganisms, such as reduction, oxidation, methylation, sulfur bonding, and transport systems. Key microbial players like *Bacillus* sp. strain DJ-1, *Thiomonas* bacteria, *Herminiimonas arsenicoxydans*, and *Sulfolobus metallicus* showcase resilience and efficacy, paving the way for biotechnological applications in water treatment, agriculture, and mining. Environmental factors significantly influence microbial bioremediation, offering a promising avenue to reduce arsenic exposure and associated health risks. Despite promises, challenges like scaling up processes and competition from other electron acceptors underscore the importance of continued research. Real-world case studies highlight success in diverse contexts, emphasizing the feasibility and efficacy of integrating microbial strategies. Regulatory considerations and globally unified policies are essential for ethical, safe, and effective implementation. The integration of microbial arsenic bioremediation with traditional methods presents a holistic, sustainable solution to arsenic contamination, advocating for a collaborative effort to harness nature's detoxpotential and safeguard human health and the environment.

Keywords: Arsenic pollution; Bioremediation; arsenicosis

#### 1. Introduction to Arsenic Contamination:

Arsenic, a natural metalloid element widely distributed in the Earth's crust, can be released into the environment through natural processes like volcanic eruptions and erosion, as well as anthropogenic activities such as mining, fossil fuel combustion, and industrial processes [26].

Exposure to arsenic, primarily through contaminated drinking water and food, constitutes a major global health concern [3]. Over 200 million people worldwide are estimated to be exposed to unsafe levels of arsenic in their drinking water. Chronic arsenic poisoning, known as arsenicosis, is associated with a variety of adverse health effects [22], including skin lesions, skin cancer, internal cancers (lung, bladder, kidney), neurological effects such as neuropathy and cognitive deficits, cardiovascular disease, diabetes, anemia, and liver damage (Fig.1).

Ingested inorganic arsenic undergoes methylation processes in the body, transforming into more toxic compounds [7]. The toxicity of arsenic depends on the form and dosage of exposure [19]. Acute high-dose exposure can lead to vomiting, diarrhea, blood disorders, and death in extreme cases. Conversely, lower chronic exposure over many years often has no immediate symptoms but can be gravely dangerous. Certain regions of the world, such as parts of South and Southeast Asia (Bangladesh, India, Vietnam), Latin America, and some areas in the United States, are especially vulnerable to high levels of arsenic in groundwater sources used for drinking, irrigation, and cooking [12]. Thiscontamination may be from natural sources but also mining or industrial activity. Mitigation strategies for arsenic exposure center on testing and monitoring drinking water sources to establish safety standards and guidelines, developing alternative water sources, and removing arsenic from water via filtration systems [21]. Agricultural techniques, including reviewing irrigation methods and arsenic uptake by plants, also require scrutiny. Overall, there is a substantial need to understand, address, and reduce arsenic contamination globally to prevent both acute toxicity and the chronic health burdens that endanger human health.

#### Microbial Arsenic Bioremediation Mechanisms:

Microorganisms have evolved various mechanisms to withstand high levels of arsenic in the environment (Fig.2). The key processes they use to detoxify and remove arsenic compounds are as follows:

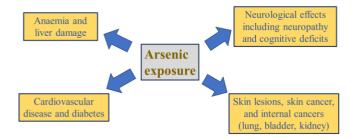


Fig.1. Effect of Arsenic on human health

**Reduction:** Many microbes reduce arsenate (AsV) to arsenite (AsIII) using a cytoplasmic arsenate reductase enzyme. Staphylococcus species, for instance, utilize the arsC gene to code for this arsenate reductase that catalyzes the reduction reaction [13]. Reduced arsenite is generally more mobile and toxic than arsenate.

**Oxidation:** Some microbes can also oxidize arsenite to arsenate using arsenite oxidase enzymes on the cell membrane [1]. This helps immobilize soluble arsenite into less soluble arsenate. Bacteria like Alcaligenes faecalis express an arsenite oxidase complex through the aoxA and aoxB genes [1].

**Methylation:** This involves adding methyl groups to inorganic arsenic using methyltransferases like the arsM gene-coded enzyme. Trivalent forms of arsenic, such as arsenite, serve as substrates for methylation, producing methylarsenite (MAsIII) and dimethylarsenite (DMAsIII). These methylated oxyanions remain toxic but are more easily exported from the cell.

**Sulfur bonding:** Microbes produce sulfur-containing molecules like glutathione and phytochelatins that can bind to and sequester arsenic intracellularly [20]. This utilizes an efflux transport system, removing arsenic from sensitive parts of the cell.

**Transport systems:** Dedicated transporter proteins, acting as arsenic pumps, actively export arsenite from the cell into the external environment. For example, E. coli utilizes ArsB and ArsAB pump proteins coded by arsenic resistance genes [25].

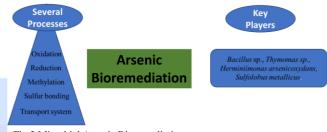
By employing various combinations of these pathways reduction, oxidation, methylation, sulfur-bonding, and membrane transport—microorganisms achieve resistance to even exceptionally high concentrations of environmental arsenic that would be toxic for most other life forms. Elucidating these microbial mechanisms is crucial for developing effective bioremediation strategies.

#### **Key Microbial Players:**

Several unique microbes have shown promising potential for arsenic bioremediation and could play a key role in cleaning arsenic-contaminated environments. *Bacillus* sp. strain DJ-1 - This Gram-positive bacterium possesses very high resistance to arsenic, able to withstand concentrations over 1000 times higher than concentrations toxic for humans. It has multiple plasmid-borne ars genes that code for arsenate reductases, arsenite oxidases, and arsenic efflux pumps [11]. DJ-1 can effectively mobilize and remove arsenic precipitates from mine tailings and industrial wastes.

*Thiomonas* bacteria - Several *Thiomonas* species can rapidly oxidize arsenite into less soluble arsenate using their arsenite oxidase enzyme. This helps immobilize and remove soluble arsenic. *T. arsenitoxydans* in particular has been studied extensively for soil/sediment remediation and arsenic removal in reactors via binding to membranes [9].

Herminiimonas arsenicoxydans - This  $\beta$ -proteobacterium is among the most resistant organisms to metals like arsenic. It possesses the ars genetic system for detoxification along with other pathways [18]. *H. arsenicoxydans* not only oxidizes As(III) into As(V) but can reduce more hazardous compounds like 4-hydroxy-3 nitrophenyl arsenic into 3nitro-4-hydroxyphenyl arsenic. This demonstrates its application in bio-treatment of industrial wastewater with complex arsenic compounds.





Sulfolobus metallicus - A thermophilic archaeon, metallicus survives in extreme high-temperature, acidic environments with high arsenic levels that would threaten most other life forms. It makes use of membrane transporters to efflux arsenic oxyanions out of cells and intothe surrounding environment [32]. This sequestration andbonding of arsenic aids bioremediation.

The unique mechanisms and high resilience of these microorganisms (Table 1) to arsenic can therefore be harnessed via metabolic engineering and bioprocessing methods to clean up arsenic from contaminated soils, industrial effluent streams, and groundwater reservoirs in an eco-friendly manner.

# **Biotechnological Applications**

Microbial arsenic bioremediation demonstrates promising applications across various industries grappling with arsenic contamination:

Bacterium	Mechanism of Arsenic	Referen ces
	Bioremediation	Ces
	Arsenate reduction,	
	arsenite oxidation,	
Bacillus sp.	arsenicmethylation and	[2,16]
	demethylation, arsenic	
	volatilization	
Pseudomonas sp.	Arsenate reduction,	[33]
	arsenite oxidation	
Burkholderia sp.	arsenite oxidation	[2]
Klebsiella sp.	arsenite oxidation	[2]
Thiobacillus sp.	Arsenite oxidation	[15]
Microbacterium	Arsenic volatilization	[27]
sp. <i>Staphylococcus</i> sp.	Arsenic adsorption	[28]
-	Arsenate	
Escherichia coli	reduction, arsenic	[8]
	transport and	
	sequestration	
Rhodopseudomo nas	Arsenate respiration	[24]
palustris	) Magnif	ivina F
Desulfitobacteriu m	Arsenate respiration	[30]
sp.		
Geobacter	Arsanata respiration	[5]
sulfurreducens	Arsenate respiration	[J]
Shewanella sp.	Arsenate respiration	[6]
Clostridium sp.	Arsenic methylation	[14]
Sulfolobus acidocaldarius	Arsenite oxidation	[31]

 Table 1: Reported bacteria involved in Arsenic bioremediation

Microbes can be employed in reactors to pre-treat influent waters with high arsenic concentrations entering water treatment plants. Through bio-oxidation and bio- reduction, these microorganisms facilitate the conversionand removal of both inorganic and complex organic arsenicals. Specialized bioreactors housing arsenic- metabolizing bacteria, such as *H. arsenicoxydans*, are particularly effective in filtering and purifying arsenic-laden water. This microbial augmentation of treatment systems proves to be more cost-effective compared to traditionalchemical treatments.

Arsenic-resistant microorganisms play a pivotal role in directly metabolizing and degrading residual arsenicals present in agricultural soils and sediments, reducing them to acceptable safety levels. Additionally, plants engineered to express bacterial arsenic resistance genes exhibit robust growth on contaminated lands unsuitable for conventional cultivation. Microbiallymediated arsenic extractions enable continued crop cultivation, avoiding the need to leave lands fallow.

Bioprocessing of mine tailings and acidic wastewaters utilizing arsenic-oxidizing microbes like *T. arsenivorans* proves effective in immobilizing solubilized arsenic. The implementation of on-site bioreactors at mining sites facilitates enhanced and safer management of arseniccontaining waste before discharge. Furthermore, solid residues can undergo bioleaching to recover other valuable metals, contributing to the economic viability of the overall process.

Moreover, the integration of microbial remediation with existing chemical and physical treatment infrastructure in a phased manner enhances efficiency for these industries. With ongoing research into microbiological mechanisms, metabolic engineering, and metagenomics, there exists substantial potential to develop more robust and scalable bioremediation processes. This progress is vital in addressing the global challenge of arsenic pollution across diverse sectors.

#### **Environmental Factors Influencing Bioremediation:**

Environmental factors, including pH, temperature, nutrients, and electron acceptors, play a crucial role in influencing the activity and efficiency of microbes engaged in metabolizing and detoxifying arsenic compounds.

pH: Microbial processes involved in the oxidation, reduction, and precipitation of arsenic compounds for detoxification exhibit high sensitivity to pH variations. Alkaline conditions favor the precipitation of insoluble arsenicals, whereas acidic conditions increase the mobility and bioavailability of ionic arsenicals. However, low pH can limit microbial growth and activity. A near-neutral circumneutral pH of 6-8 proves to be optimal for many arsenic-resistant bacteria and archaea.

Temperature: Microbes exhibit diverse temperature tolerances, ranging from mesophilic to thermophilic and psychrophilic arsenic metabolizers based on their habitat. Moderately warm temperatures often accelerate enzyme kinetics, while extremely high heat can damage transporters and proteins. Low cold temperatures reduce membrane fluidity and transport functions. Therefore, optimizing temperature is vital to ensuring microbial efficiency.

Nutrients: Phosphate limitation hampers biosynthesis and restricts arsenate reduction by microbes. Nitrogen sources like ammonium can inhibit microbial oxidation of As(III). Conversely, certain organic nutrients derived from plant roots stimulate As oxidation by soil microbes.

Electron acceptors: The activity of dissimilatory arsenatereducing bacteria relies on the availability of electron acceptors, such as nitrates, sulfates, iron, and bicarbonate ions. As a result, the geochemistry and mineral composition of contaminated matrices significantly impact arsenic redox transformations.

Understanding how these influential environmental variables modulate microbial activities provides valuable insights for engineering field applications of bioreactors, improving enzymatic processes, and developing site-specific bioremediation approaches tailored to native conditions. Nutrient amendments and the optimization of physicochemical parameters will contribute to maximizing arsenic mobilization and detoxification by microorganisms.

#### Human Health Implications:

The advancement of microbial arsenic bioremediation techniques holds significant promise for improving human health outcomes by mitigating arsenic exposures and associated morbidity and mortality burdens on a global scale.

These innovative biotechnologies work by metabolizing and removing environmental arsenic contamination, directly reducing human contact with arsenic through critical pathways such as drinking water and the food chain. For instance, the introduction of arsenite-oxidizing bacteria in water treatment systems ensures that delivered water contains extremely minimal residual arsenic, well within safety limits, in stark contrast to the dangerously high concentrations observed before such interventions.

The resulting reduction in arsenic exposures carries extensive implications for human health in both short and long-term scenarios. This includes a substantial decrease in cases of acute arsenical poisoning, marked by symptoms like diarrhea, vomiting, seizures, and shock syndromes, resulting in a rapid decline in mortality rates. Additionally, there is a lower incidence of characteristic arsenic dermatosis and keratosis skin lesions, which have the potential to transform into non-melanoma cancers, such as

Bowen's disease. Furthermore, the implementation of microbial bioremediation techniques contributes to diminished rates of bladder, kidney, liver, and lung cancers in populations chronically exposed to arsenic over extended periods, thereby lowering both cancer morbidity and associated mortalities. The positive impact extends to improved clinical outcomes and a reduced prevalence of cardiovascular disease, hematologic disorders like anemia, diabetes, obstetric complications, and neurological effects linked to arsenic exposure. This comprehensive reduction in arsenic-related health risks underscores the transformative potential of microbial bioremediation, not only in preventing acute poisonings but also in mitigating the long-term burden of various diseases associated with chronic arsenic exposure.

Through these broad epidemiological and toxicological benefits spanning mortality, cancer outcomes, and noncommunicable diseases, the application of microbial bioremediation techniques significantly enhances public health prospects globally while concurrently reducing environmental arsenic contamination. The integration of these bio-approaches with traditional water treatment infrastructure and improved medical care access further potentiate these positive human health effects.

#### **Challenges and Future Directions**

potential, Despite microbial its strong arsenic bioremediation faces several challenges that currently limit its more widespread and efficient applications, necessitating further research. One primary challenge is the insufficient understanding of environmental microbiology and geochemistry. To overcome this, more in-depth research focusing on native microbial communities in contaminated sites and the complex aqueous geochemistry governing arsenic speciation is essential. This will enable the development of tailored, site-specific solutions. Another significant obstacle lies in scaling up laboratory bioprocesses to field sites. Questions persist regarding the industrial-scale propagation of arsenic-metabolizing cultures and the application of genetic engineering advances to bioreactors for long-term operational viability. Additionally, competition from other electron acceptors, such as nitrates, proves to be a noteworthy challenge. In the field, these alternate contenders inhibit microbial arsenic respiration pathways more significantly compared to controlled lab settings. Defining amendments and optimization regimes to effectively address this competition is crucial for successful bioremediation. Furthermore, comprehensive cost-benefit analyses for technology adoption are imperative. Demonstrating favorable economic incentives, coupled with performance metrics, will play a vital role in encouraging greater industry investment in bioremediation approaches over standard chemical treatments. Addressing these challenges through multi-disciplinary research efforts will be pivotal in unlocking the full potential of microbial arsenic bioremediation, making it more efficient and applicable on a broader scale.

Future avenues to focus on include elucidating novel microbial pathways and enzymes for arsenic conversion using meta-omics techniques, developing Metal-Organic Frameworks (MOFs) and nanomaterials to enhance bacterial degradation, creating arsenic-specific biosensors for rapid diagnostics, assessing field-scale viability in randomized control trials, and integrating ecological engineering principles for sustainable bioremediation that maintains ecosystem health while detoxifying arsenic pollution. Addressing these challenges through multi-disciplinary research will serve to unlock the immense potential of microbes for efficient, eco-friendly arsenic bioremediation.

#### **Case Studies**

Promising real-world applications of microbial bioremediation have effectively addressed arsenic contamination in various contexts. In Cambodian groundwater treatment systems, arsenic biosand filters augmented with iron-oxidizing bacteria successfully lowered arsenic levels in drinking water to meet WHO guidelines. Over a 6-month project across 25 households, arsenic levels plummeted from an alarming 600 ppb initially to a mere 2 ppb. In a US mine site rehabilitation effort, bioreactors employing Sulfolobus acidocaldarius remediated over 180 million liters of acidic, metal-rich water from the Iron Mountain mine over a decade. The bioremediation process achieved a remarkable 99% arsenic removal efficiency through bio-oxidation and immobilization. A bioremediation study in Vietnam focused on model aquifer columns inoculated with native arsenic- metabolizing strains Desulfotomaculum and Clostridium. This approach led to an over 80% reduction in aqueous arsenic through sorption and precipitation, demonstrating the successful scaling up of lab studies. Lastly, in West Bengal, farmer field schools implemented participatory action projects that trained farmers in using on-site bioremediation enriched with nutrients to treat arsenic- affected paddy soils. The resulting yield improvements prompted community adoption, showcasing the socioeconomic viability of microbial bioremediation in real- world scenarios. These cases underscore the potential of microbial bioremediation as a practical and impactful solution for arsenic contamination across diverse environments. These cases highlight key success factors like process integration with existing infrastructure, involvement of end-user communities, controlled lab validation prior to scaling up, and examining both technological efficacy and cost-benefit impact.

However, long-term monitoring, evaluating sustainability metrics beyond narrow project durations, and comparing to alternative remediation approaches are vital to guide evidence-based policy. Synthesizing more such field demonstrations and constructive analyses will serve to refine microbial bioremediation approaches for managing the global arsenic challenge.

## **Regulatory Considerations:**

Currently, there are no comprehensive globally unified policies or regulatory guidelines focused specifically on microbial arsenic bioremediation. However, we can extrapolate some frameworks from related environmental and biotechnology policies to ensure safe, ethical, and effective implementation. Firstly, the environmental release and deployment of engineered arsenic-remediating microorganisms will need to conform to nationally accredited biosafety rules and procedures under bodies like the NIH and CDC in the United States or Gene Technology Regulator in Australia. Following established protocols for risk assessment of environmental exposure, hazard characterization, containment mechanisms before release can mitigate uncertainties. Additionally. relevant environmental protection agencies like the EPA or EEA should provide science-based guidance on acceptable total arsenic limits across different matrices like soil, water, that bioremediation efforts cropsmust achieve. Monitoring protocols for sampling and SVD analysis should feature standardization. Conflict of interest clauses are also integral to evaluation processes.

From a social perspective, the Aarhus convention provisions on access to information and public participation should frame bioremediation projects for communal ownership. Policies should also promote fair access to this technology alongside traditional water treatment plants in marginalized areas to ensure environmental justice. Frameworks around technology transfer, IPs, and profit sharing need reviewal to facilitate rapid innovation diffusion from lab research to industrial use in developing countries bearing the highest arsenic burden. Incentives for green chemistry solutions over chemical methods are also key.

Overall, a hybrid approach balancing biosafety, consumer interests, conservation needs, and community priorities through evidence-based policies will allow microbial arsenic bioremediation to deliver on its public health promises sustainably.

# Integration with Traditional Remediation Methods:

Microbial arsenic bioremediation provides an eco- friendly, low-cost alternative that can synergistically integrate with conventional physiochemical remediation methods for contaminated site cleanup and water treatment.

Various integration pathways enhance the effectiveness of microbial arsenic bioremediation in different contexts:

One approach involves the use of bioreactors inoculated with arsenic-metabolizing bacteria, such as Rhodopseudomonas palustris, as a polishing step after chemical processes like coagulation, lime softening, and ion exchange. This integration significantly improves overall arsenic removal efficiency, ensuring compliance with stringent limits [17]. A phased approach proves effective by employing microbial bioprocessing to immobilize and separate bulk arsenic from waste streams initially. Following this, techniques like membrane filtration or adsorption can be applied more judiciously at lower arsenic concentrations. On-site microbial bioremediation of solid wastes represents another integration strategy. This method helps concentrate arsenic into leachates, which can then undergo chemical precipitation or be directed to centralized plants for conventional treatment more easily (Taran et al., 2019). Utilizing byproducts or wastes from other remediation processes can nourish and stimulate microbial metabolism. For instance, biosludge ash from incinerators can serve as bioavailable phosphate sources, enhancing microbial activity [29]. Additionally, integrating ecological engineering principles, such as constructed treatment wetlands harboring microbial biofilms, offers a low-energy and low-cost complementary solution, particularly suitable for small communities [4]. These diverse integration pathways showcase the adaptability and versatility of microbial bioremediation in arsenic-contaminated environments.

Overall, strategically applying microbial and physiochemical remediation in tandem by exploiting their distinct strengths can lead to synergies in performance.

Policy incentives to install such integrated systems, especially in remote regions lacking centralized water infrastructure can help drive more sustainable innovation.

#### Conclusion

In conclusion, the review paper "Unveiling Nature's Detox: Microbial Arsenic Bioremediation for Human Health" delves into the multifaceted issue of arsenic contamination, emphasizing the global health threat it poses. From the insidious chronic effects of arsenicosis to the acute dangers of high-dose exposure, the paper underscores the urgency of addressing this pervasive problem.

The exploration of microbial arsenic bioremediation mechanisms sheds light on the ingenious strategies employed by microorganisms to detoxify arsenic. Reduction, oxidation, methylation, sulfur bonding, and transport systems are the tools these microorganisms use to thrive in high-arsenic environments, offering a foundation for potential biotechnological applications.

Key microbial players such as *Bacillus sp.* strain DJ-1, *Thiomonas* bacteria, *Herminiimonas arsenicoxydans*, and *Sulfolobus metallicus* demonstrate the remarkable resilience and efficacy of certain microorganisms in arseniccontaminated settings. Their unique abilities pave the way for innovative biotechnological applications across water treatment, agriculture, and the mining industry.

Environmental factors significantly influence microbial bioremediation, with pH, temperature, nutrients, and electron acceptors playing crucial roles. Understanding these factors provides valuable insights for optimizing bioremediation processes in diverse environmental conditions.

The implications for human health are profound, with microbial arsenic bioremediation offering a promising avenue to reduce arsenic exposure and associated health risks. By mitigating the risk of arsenic-induced diseases and improving public health outcomes, these biotechnological interventions hold the potential to make a substantial impact globally.

Despite the promises, challenges such as scaling up processes, competition from other electron acceptors, and the need for cost-effective solutions underscore the importance of continued research. Future directions point toward a more comprehensive understanding of microbial pathways, advancements in genetic engineering, and the development of sustainable bioremediation processes.

Real-world case studies highlight the success of microbial bioremediation in diverse contexts, from Cambodian groundwater treatment systems to US mine site rehabilitation. These cases emphasize the feasibility and efficacy of integrating microbial strategies into existing environmental management practices. Regulatory considerations and the need for globally unified policies underscore the importance of ethical, safe, and effective implementation of microbial arsenic bioremediation. Balancing biosafety, environmental protection, and community engagement is paramount for the success of these technologies.

The integration of microbial arsenic bioremediation with traditional methods offers a holistic approach to environmental cleanup. Whether in water treatment plants, agriculture, or mining, the synergistic application of microbial and physiochemical remediation methods presents a sustainable and efficient solution to the complex challenge of arsenic contamination.

In essence, the review paper advocates for a comprehensive and collaborative effort to unveil nature's detox potential through microbial arsenic bioremediation.By harnessing the power of microorganisms, we stand at the forefront of a transformative approach to safeguarding human health and the environment from the pervasive threat of arsenic contamination.

# Acknowledgments

Review work is made during the research work of the first author. Dr. Arnab Ganguli helps a lot to structure the article in a manuscript format. Techno India University, West Bengal provides the infrastructures as well as environment to grow research mentality.

#### References

- Bahar, M. M., Megharaj, M., & Naidu, R. (2016). Oxidation of arsenite to arsenate in growth medium and groundwater using a novel arsenite-oxidizing diazotrophic bacterium isolated from soil. *International Biodeterioration & Biodegradation*, 106, 178-182.
- [2] Chakraborty, A., & Islam, E. (2018). Temporal dynamics of total and free-living nitrogen-fixing bacterial community abundance and structure in soil with and without history of arsenic contamination during a rice growing season. *Environmental Science and Pollution Research*, 25, 4951-4962.
- [3] Ciesielczuk, T., Rosik-Dulewska, C., Poluszyńska, J., Ślęzak, E., & Łuczak, K. (2018). Ashes from sewage sludge and bottom sediments as a source of bioavailable phosphorus. *Journal of Ecological Engineering*, 19(4).
- [4] Dang, Y., Walker, D. J., Vautour, K. E., Dixon, S., & Holmes, D. E. (2017). Arsenic detoxification by Geobacter species. *Applied and Environmental Microbiology*, 83(4), e02689-16.
- [5] Darma, A., Yang, J., Zandi, P., Liu, J., Możdżeń, K., Xia, X., ... & Schnug, E. (2022). Significance of Shewanella Species for the Phytoavailability and Toxicity of Arsenic—A Review. *Biology*, 11(3), 472.
- [6] Drobna, Z., Styblo, M., & Thomas, D. J. (2009). An overview of arsenic metabolism and toxicity. *Current protocols in toxicology*, 42(1), 4-31.
- [7] Drobna, Z., Styblo, M., & Thomas, D. J. (2009). An overview of arsenic metabolism and toxicity. *Current protocols in toxicology*, 42(1), 4-31.
- [8] Edmundson, M. C., & Horsfall, L. (2015). Construction of a modular arsenic-resistance operon in E. coli and the production of arsenic nanoparticles. *Frontiers in Bioengineering and Biotechnology*, 3, 160.
- [9] Hovasse, A., Bruneel, O., Casiot, C., Desoeuvre, A., Farasin, J., Hery, M., ... & Arsène-Ploetze, F. (2016). Spatio-temporal detection of the Thiomonas population and the Thiomonas arsenite oxidase involved in natural arsenite attenuation processes in the Carnoulès acid mine drainage. *Frontiers in Cell and Developmental Biology*, 4, 3.

- [10] Huertas, E., Folch, M., & Salgot, M. (2007). Wastewater reclamation through a combination of natural systems (infiltrationpercolation and constructed wetlands): a solution for small communities. *Water science and technology*, 55(7), 143-148.
- [11] Huertas, E., Folch, M., & Salgot, M. (2007). Wastewater reclamation through a combination of natural systems (infiltration-percolation and constructed wetlands): a solution for small communities. *Water science and technology*, 55(7), 143-148.
- [12] Kim, K. W., Chanpiwat, P., Hanh, H. T., Phan, K., & Sthiannopkao, S. (2011). Arsenic geochemistry of groundwater in Southeast Asia. *Frontiers of medicine*, 5, 420-433.
- [13] Kumari, N., & Jagadevan, S. (2016). Genetic identification of arsenate reductase and arsenite oxidase in redox transformations carried out by arsenic metabolising prokaryotes–A comprehensive review. *Chemosphere*, 163, 400-412. *Geomicrobiol. J.* 33, 3–4 (March 2016), 185–193.<u>https://doi.org/10.1080/01490451.2015.1052117</u>
- [14] Li, B., Deng, C., Zhang, D., Pan, X., Al-misned, F. A., & Mortuza, M. G. (2016). Bioremediation of nitrate-and arsenic-contaminated groundwater using nitrate-dependent Fe (II) oxidizing Clostridium sp. strain pxl2. *Geomicrobiology Journal*, 33(3-4), 185-193.
- [15] Li, Y., Guo, L., Yang, R., Yang, Z., Zhang, H., Li, Q., ... & Sun, W. (2023). Thiobacillus spp. and Anaeromyxobacter spp. mediate arsenite oxidation-dependent biological nitrogen fixation in two contrasting types of arsenic-contaminated soils. *Journal of hazardous materials*, 443, 130220.
- [16] Magar, L. B., Rayamajhee, B., Khadka, S., Karki, G., Thapa, A., Yasir, M., ... & Poudel, P. (2022). Detection of Bacillus Species with Arsenic Resistance and Plant Growth Promoting Efficacy from Agricultural Soils of Nepal. *Scientifica*, 2022.
- [17] Mohsin, H., Asif, A., & Rehman, Y. (2019). Anoxic growth optimization for metal respiration and photobiological hydrogen production by arsenic-resistant Rhodopseudomonas and Rhodobacter species. *Journal of basic microbiology*, 59(12), 1208-1216.
- [18] Muller, D., Simeonova, D. D., Riegel, P., Mangenot, S., Koechler, S., Lievremont, D., ... & Lett, M. C. (2006). Herminiimonas arsenicoxydans sp. nov., a metalloresistant bacterium. *International journal of systematic and evolutionary microbiology*, 56(8), 1765-1769.
- [19] Ng, J. C. (2005). Environmental contamination of arsenic and its toxicological impact on humans. *Environmental Chemistry*, 2(3), 146-160.
- [20] Pawlik-Skowrońska, B., Pirszel, J., Kalinowska, R., & Skowroński, T. (2004). Arsenic availability, toxicity and direct role of GSH and phytochelatins in As detoxification in the green alga Stichococcus bacillaris. *Aquatic Toxicology*, 70(3), 201-212.
- [21] Pearson, M., Jones-Hughes, T., Whear, R., Cooper, C., Peters, J., Evans, E. H., & Depledge, M. (2011). Are interventions to reduce the impact of arsenic contamination of groundwater on human health in developing countries effective?: a systematic review protocol. *Environmental Evidence*, 1, 1-7.
- [22] Rahman, M. M., Ng, J. C., & Naidu, R. (2009). Chronic exposure of arsenic via drinking water and its adverse health impacts on humans. *Environmental geochemistry and health*, 31, 189-200.
- [23] Rhine, E. D., Chadhain, S. N., Zylstra, G. J., & Young, L. Y. (2007). The arsenite oxidase genes (aroAB) in novel chemoautotrophic arsenite oxidizers. *Biochemical and biophysical research communications*, 354(3), 662-667.
- [24] Sabki, M. H., Ong, P. Y., Lee, C. T., Ibrahim, N., Van Fan, Y., & Klemeš, J. J. (2021). The Potential of Rhodopseudomonas Palustris as a Bio-Fertiliser for Sustainable Agriculture. *Chemical Engineering Transactions*, 88, 457-462.
- [25] Saltikov, C. W., & Olson, B. H. (2002). Homology of Escherichia coli R773 arsA, arsB, and arsC genes in arsenic-resistant bacteria isolated from raw sewage and arsenic-enriched creek waters. *Applied and Environmental Microbiology*, 68(1), 280-288.
- [26] Schreiber, M. E., & Cozzarelli, I. M. (2021). Arsenic release to the environment from hydrocarbon production, storage, transportation, use and waste management. *Journal of Hazardous Materials*, 411, 125013.

- [27] Sher, S., Hussain, S. Z., Cheema, M. T., Hussain, A., & Rehman, A. (2022). Efficient removal potential of Microbacterium sp. strain 1S1 against arsenite isolated from polluted environment. *Journal of King Saud University-Science*, 34(5), 102066.
- [28] Srivastava, S., Verma, P. C., Singh, A., Mishra, M., Singh, N., Sharma, N., & Singh, N. (2012). Isolation and characterization of Staphylococcus sp. strain NBRIEAG-8 from arsenic contaminated site of West Bengal. *Applied microbiology and biotechnology*, 95, 1275-1291.
- [29] Taran, M., Fateh, R., Rezaei, S., & Gholi, M. K. (2019). Isolation of arsenic accumulating bacteria from garbage leachates for possible application in bioremediation. *Iranian Journal of Microbiology*, 11(1), 60.
- [30] Villemur, R., Lanthier, M., Beaudet, R., & Lépine, F. (2006). The desulfitobacterium genus. *FEMS microbiology reviews*, 30(5), 706-733.
- [31] Yin, S., Zhang, X., Yin, H., & Zhang, X. (2022). Current knowledge on molecular mechanisms of microorganism-mediated bioremediation for arsenic contamination: A review. *Microbiological Research*, 258, 126990.
- [32] Zangi, R., & Filella, M. (2012). Transport routes of metalloids into and out of the cell: a review of the current knowledge. *Chemico-Biological Interactions*, 197(1), 47-57.
- [33] Zhang, Z., Yin, N., Cai, X., Wang, Z., & Cui, Y. (2016). Arsenic redox transformation by Pseudomonas sp. HN-2 isolated from arseniccontaminated soil in Hunan, China. *Journal of Environmental Sciences*, 47, 165-173.

Facts for Solutions