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Abiotic Stress Tolerance in Plants: Insights into the Functions of Glutathione Peroxidases

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Abstract

Years of evolution and numerous adaptations have developed plants to be a unique eukaryotic multicellular organism which is independent in nature. One of such miraculous features of plant is its inbuilt defense mechanism which had protected itself from any extreme condition. Unfortunately, we human and our action had over exploited every natural resource to such an extent that the "Mother Earth" which gave us life, are undergoing drastic changes hampering its actual ambience. One of the most recent concerning issues that had made scientist distressed is the rapid change of climatic condition of nature, which therefore followed by increase in rate of abiotic stress. Due to this increase abiotic stresses, high rate of ROS production takes place in plant which create an imbalance in its homeostasis. Reactive Oxygen Species (ROS) are highly reactive and toxic which effects the biosynthesis of chlorophyll, photosynthetic capacity, and carbohydrate, protein, lipid, and antioxidant enzyme activities. ROS is the byproduct of cellular metabolism of plant cell which triggers the stress signal transduction activating the defense mechanism, that is synthesis of antioxidants which maintains the level of this reactive species. The activity of several types of enzymatic and non-enzymatic antioxidant has greatly contributed on the defense mechanism of plants helping it to withstand the adverse condition of the environment. In this article, we will be assessing the process of how the plants physiological and biochemical changes take play on exposure to these abiotic stresses and how antioxidant in cell combat these changes for plant to survive.

Keywords: ROS, Plant Abiotic Stress Physiology, GPX, Antioxidant defense mechanism, RNS, Lipid Peroxidation

1. INTRODUCTION

Plant belonging to kingdom Plantae are eukaryotic organism, having an autotrophic mode of nutrient. It undergoes the process of photosynthesis in which CO2 and water are combined in the presence of sunlight trapped by Mg2+ containing green pigment, known as chlorophyll in order to synthesize its energy from inorganic compound [1]. According to the above mention fact it proves that plants are self-dependent in synthesizing their own food, it is also self-sufficient in facing and adapting itself according to the adverse environmental condition occurred due to naturally or man-made to

certain extent. The adverse environmental conditions also known as environmental stress occurring mainly due to sudden climatic changes which is consequences of sessile human lifestyle. When environmental stress occurs due to biotic factor then it is known as biotic stress and when occurs due to abiotic factor it is known as abiotic stress. Abiotic stress has now become one of the most extensive studies for researchers and its effect on plants and mainly on field crops [2]. It's various type such as radiation UV A and UV B, water (drought and flood), salinity, temperature (high and low), chemical factors (heavy metal and pH), nutrient (deficiency and excess), light (high and low) intensity, gaseous

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pollutant (ozone, sulfuric gas), mechanical factor and anaerobic has garnered quite a lot attention of researchers [2,6]. This leads to the discovery of all type of coping mechanism adapted by the plants to deal these stresses and to sustain normal growth under such condition. Sometimes combination of these stresses brings unique cellular level modification that cannot be expected from individual stressors [6]. These modification results in biochemical and physiological changes in plant, generating certain secondary metabolites which induces stress signals which then help the plant to produce stress signaling pathways in response to it. These pathways help to maintain homeostasis and acclimating itself to the modified conditions. These changes are also expressed in their genes [5], inherited by their progeny hence evolving in order to survive in these adverse changes. Major of this abiotic stress researches are done in laboratory, under controlled conditions which may vary with the condition that actually occurs in the field [3]. Environmental stress has become a concerning issue among scientist due to its detrimental effect on crop productivity and its development, majorly to meet the demands of enhanced food requirement (Table 1).

Abiotic	Key Effects		
Stress	Magniyng		
Drought	Reduced cell division, smaller leaf		
	size, nutrient uptake hindrance, water		
	loss.		
Salinity	Ion toxicity, disrupted		
	photosynthesis, membrane disorders,		
	oxidative stress.		
Temperature	Delayed or early flowering,		
	reproductive phase disruptions.		
Heavy	ROS overproduction, biomolecule		
Metals	inactivation, oxidative stress.		

As by the word "abiotic" we understand something that is not obtain from living organism. Abiotic factors are physical rather than biological. Abiotic factors in environment are all the natural resources such as sunlight, water, temperature, soil, wind, humidity etc. Human's overexploitation of these natural resources had lead to the drastic changes in natural course of natural resources resulting extreme conditions known as "stress" or more specifically "abiotic stress". Plant getting exposed to such abiotic stresses have independently developed its coping mechanism by making changes in its cellular metabolism.

Plants exposed to abiotic stress are found to have an increase in amount of ROS (Reactive Oxygen Species) which are highly reactive and toxic, affecting the biosynthesis of chlorophyll, photosynthetic process, and carbohydrate, protein, lipid and antioxidant enzyme activities [8]. Plant use molecular oxygen for its cellular metabolism. Reactive Oxygen species also known as ROS are the byproduct mitochondrial of oxidative phosphorylation and also by cellular response to xenobiotics, cytokines, and bacterial invasion. Reactive oxygen species such as superoxide anion (O_2) , hydrogen peroxide (H_2O_2) , and hydroxyl radical (HO•), are of radical and non-radical oxygen species formed by the partial reduction of oxygen [34] in sites such as chlorophyll, mitochondria both are double membraned subcellular organelles [40] and Peroxisomes is single membraned subcellular organelles [41]. This increase in accumulation of ROS in cell disturb the homeostasis and ion distribution [40,42] which increases the oxidative stress in plant [34]. ROS-mediated oxidative stress causes damage of nucleic acids, proteins, and lipids involves in carcinogenesis [35], neurodegeneration [36,37], atherosclerosis [34], diabetes [38], and aging [39]. So plants are fully evolved to regulate all this overwhelming production of ROS and to thereby reduces the toxic effect of high ROS by the production of antioxidants. Antioxidants are biomolecules that reduces, repairs or prevent the effect by inhibiting the oxidation of oxygen species present in ROS (e.g. O_2^- , H_2O_2 , OH^- , O_2) [4]. They are first line of defense against the harmful effects caused by free radicals [33,43,44,45]. Plant antioxidants have an important role in plant development and has a wide range of mechanism and function [40]. Plant antioxidant detoxify in three major ways i) antioxidants that forage produced ROS ii) antioxidant that inhibit ROS production iii) antioxidant that cures or alters the damage caused by ROS [7]. It does not completely eradicate oxidants

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but rather maintains an optimum level [46]. Antioxidant defense system comprises of two types :i)Enzymatic antioxidant

ii) non-enzymatic antioxidant (Table 2).

Table 2: Antioxidant Defense Mechanisms inPlants

Antioxidant	Key Enzymes	Function
Туре		
Enzymatic	SOD, CAT,	Neutralizes
Antioxidants	GPX, APX	ROS by
		converting
		them to non-
		toxic forms.
Non-Enzymatic	Ascorbic acid,	Maintains
Antioxidants	Glutathione,	redox balance,
	Phenolic	scavenges
	compounds,	ROS.
	Flavonoids	

- → Enzymatic antioxidant superoxide dismutase (SOD), catalase (CAT), peroxidases (POX), glutathione peroxidase (GPX), glutathione reductase (GR), glutathione S-transferases (GST), ascorbate peroxidase (APX), monodehydroascorbate reductase (MD-HAR) and dehydroascorbate reductase (DHAR) [47-49].
- Non-enzymatic antioxidant ascorbic acid (AA), glutathione (GSH), phenolic compounds, alkaloids, flavonoids, carotenoids, free amino acids and α-tocopherols [47-49].

As this work primary focuses on **GLUTATHIONE PEROXIDASE** which falls under the category of PEROXIDASE enzyme, so we will be discussing about the PEROXIDASE enzymes.

2. PEROXIDASE(POX):-

Peroxidase is a hydrogen peroxide (H_2O_2) decomposing enzyme accompanying oxidation of various phenolic and non-phenolic substrates. Its application as an industrial enzyme for medicinal, immunological, biotechnological aspect, bioremediation, textile etc.[68] Its various use makes it one of the primary enzyme

Peroxidase	Function	Examples
Туре		
Heme	Catalyzes	Myeloperoxidase
Peroxidase	H2O2	(MPO), Eosinophil
	decomposition;	Peroxidase (EPO).
	innate	
	immunity.	
Non-Heme	Reduces	Thiol Peroxidase,
Peroxidase	hydroperoxides	Alkylhydroperoxidase.
	and detoxifies	
	lipid peroxides.	
Class III	Oxidoreduction	Cytochrome c
Plant-	of H2O2; stress	Peroxidase, Ascorbate
Specific	response.	Peroxidase.

Table 3: Characteristics of Peroxidase Enzymes

Peroxidase is categorized into two types:

- A) Heme peroxidase B) Non-Heme peroxidase [68]
- A) Heme peroxidase is further divided into two:
 - Peroxidase-Cyclooxygenase Superfamily (PCOXS) – is animal specific peroxidase and takes part in innate immunity, defense mechanism etc. E.g.- Myeloperoxidase (MPO), Eosinophil peroxidase (EPO), Lactoperoxidase (LPO), Thyroid peroxidase (TPO)
 - Peroxidase-Catalase Superfamily (PCATS)
 is a plant specific peroxidase which is further subdivided into following three classes [53]:-
 - Class I plant peroxidase involves intracellular enzymes in plants, bacteria and yeast such as
 - Cytochrome c peroxidase {EC 1.11.1.5}
 - Bacterial catalase-peroxidase {EC 1.11.1.6}
 - Ascorbate peroxidase {EC 1.11.1.11}
 - Class II plant peroxidase extracellular peroxidase from fungi such as
 - Lignin peroxidase {EC 1.11.1.14}
 - Mn²⁺-dependent peroxidase {EC 1.11.1.13}

- Class III plant peroxidase Peroxidases {EC 1.11.1.7}
- B) Non-Heme peroxidase is further divided into:
 - 1) Thiol Peroxidase
 - 2) Halo-Peroxidase
 - 3) Alkylhydroperoxidase
 - 4) NADH peroxidase

Peroxidases enzyme (POX) {EC number 1.11.1.7} is a Class III plant-specific peroxidase whose function is to catalyze oxidoreduction of H_2O_2 . It is usually stable at high temperature and its activity is determined by using simple chromogenic reactions, which makes it an ideal enzyme for study of protein structure, enzyme reaction and enzyme functions for any practical application [51] such as immunoassays, diagnostic assays and industrial enzymatic reaction [50]. Peroxidase is a heme-containing glycoprotein which encode a large multigene family in plant [51].



In spite of GLUTAHTHIONE PEROXIDASE {EC 1.11.1.9} falls under the animal peroxidase super-family, it is noticed that its activity is found in plants and various other cDNAs encoding its homologs [52,56,57]. So, focusing on actual topic of this review paper that is GLUTATHIONE PEROXIDASE, here we will be discussing about its

characteristic and its tolerance against certain abiotic stress.

2.1. GLUTATHIONE PEROXIDASE:-

Thiol peroxidase is a non-heme peroxidase which includes thioredoxin peroxidases or peroxiredoxins (Prxs) and Glutathione peroxidases (GPXs). On the basis of their amino acid sequence, substrate specificities, and subcellular localization, GPX is divided into five classes [69]. Glutathione peroxidase is chief compound in gluthathione-ascorbate cycle which reduce H₂O₂ by oxidizing reduced glutathione (GSH) to disulfide form (GSSH) [70,71]. Α conserved catalytic cysteine near the N-terminus of these protein is known as peroxidatic cysteine (Cys_p-S) whose function is to reduce hydroperoxides and peroxynitrites. First the Cys- residue is transformed into a sulfenic acid (CysP-SOH) when exposed to peroxides. The main difference between the different classes is the mechanism of regeneration of the CysP-SOH, which can be reduced directly (1-Cys mechanism) or by involving a second, so called resolving Cys residue (CysR-SH), which condenses with the sulfenic acid to form a disulfide (2-Cys catalytic cycle). The 2-Cys disulfide is reduced by thioredoxin - a low-molecular weight protein with two vicinal Cys residue - or by glutathione (reduced form GSH, -glu-cys-gly) [54,55]. As said before GPX is found in mammalian cell, but noticed its activity in plant cell [56,57] and has function such as detoxifying lipid hydroperoxides and other reactive molecules in different species undergoing several stress[59-63]. Plant GPX gene showing homology with animal GPX gene ((GPx4/PHGPX enzyme) are isolated from plant[54]. Plant GPXs are found to be in monomeric form [64] except for the poplar GPX5, which showed an unique dimerization pattern mainly depending on hydrophobic contacts and was able to interact with Cd2+ ions [65]. Glutathione are considered as actual thioredoxin peroxidase [66]. Studies suggest that plant GPX is considered to be more effective in reducing peroxides different from H₂O₂ such as organic hydroperoxides and lipid peroxides [67].

Table 4: GPX Activity under Different Abiotic Stresses

Abiotic	Observed GPX Activity
Stress	
Drought	High activity in guard cells,
	decreased glutathione pool in rice
	seedlings, negligible activity in
	barley root tips.
Salinity	GPX activity correlates with
	maintaining homeostasis in
	chickpea and mangroves, high
	activity observed in GPX3 rice
	plants.
Temperature	Moderate heat stress increases
	GPX activity in apple leaves;
	activity peaks at 4 hours under
	high temperatures.
Heavy Metals	Increased GPX activity under Cd
	and Hg stress; negligible activity
	for Co exposure.

As for now we have a thorough knowledge about the enzyme Glutathione peroxidase (Table 4), hence we will be discussing about how different abiotic stress affecting the environment and how GPX is able combat those abiotic stresses:

Drought stress -Crops exposed to severe climatic condition, had affected its growth and development, which then hinders the productivity of plants [13,15,17]. Drought stress is considered one of most detrimental environmental stress than any other stress [14,15]. Reduction in rainfall with higher transpiration rate leads to agricultural drought [13]. Lack of water in soil, results in reduced rate of cell division, expansion of leaf size, stem elongation and root proliferation and deficiency of certain nutrients [15,18]. Water deficiency enhances abscisic acid (ABA) biosynthesis, reducing stomatal conductance to minimize transpirational losses [15,19]. ABA signaling pathway is activated by the drought stress response[5,72]. Studies of chemical genetics and protein-interaction recognizes the PYR/ PYL/RCAR, proteins START domain as receptors for ABA[73,74].

In Arabidopsis thaliana, studies suggest that Glutathione peroxidase 3(ATGPX3) shows excess water loss, high sensitivity to H O and excess accumulation in guard cell [69]. In rice seedling grown for 20 days when exposed to drought stress hand shown a 20-38% decrease in glutathione concentration from total glutathione pool [75]. In barley root tip, negligible GPX activity is found against drought stress [76].

Salinity stress - Soil salinity is a worldwide problem affecting approximately 20% of cultivating land, reducing growth and development of plant in field [21,22], becoming one of serious limiting factor for agricultural basis [8]. Soil with high salinity contains high concentration of soluble salt and exchangeable sodium on surface [8,23], which then affects the root system of the plant[8]. Salinity not only affects the roots in plant it also affects in many other ways such as the alternation metabolic processes, membrane disorders, irregular cell division, reduction in photosynthetic activity, protein synthesis, increase in plant toxicity and enzymatic disorder [8,24]. Plants undergoing salinity stress takes up excess amount of Na⁺ and Cl⁻ ions, hence increasing accumulation of ion in tissues of plant, leading to oxidative stress [8,25]. Such accumulation result in increased toxicity effecting by inhibiting protein formation, photosynthesis and susceptible enzymes [8,25]. Plant mechanism to respond such stresses Na⁺/K⁺ homeostasis and Na⁺ exclusion [26]. Excessive accumulation of Na⁺ and Cl⁻ in plants results in overproduction of ROS [24].

In barley root tip, no activity of GPX is visible [76]. When chickpea (Cicer arietinum) cultivation is taken place with different level of saline water, there is increase hydrogen peroxide and lipid peroxidation, treatment with Glutathione resulted in decrease level of hydrogen peroxide and lipid peroxidation, thereby maintaining homeostasis [77]. GPX3s rice plants are shown to be sensitive towards salinity stress, presences of GPX3 helps to withstand the stress [78]. While working with mangroves, scientists had found a direct correlation between the concentration of Peroxidase enzyme and Salinity [80, 81]. They found out that for majority of the mangrove plant species the concentration of peroxidase increased along the salinity gradient up to a certain threshold level, beyond which it declined showing the inability of the plant to sustain any further salinity.

Temperature stress - Among many abiotic factor, optimum temperature is one of the crucial factors that helps to sustain life on earth. Extreme heat and cold temperatures have awful influence on all stages of plant growth and development but mainly on its reproductive stages [8]. According to the Intergovernmental Panel on Climate Change (IPCC), growth of plant will be challenged with warmer environment as the average surface temperature will increase 2.0-4.5°C by the end of this century [8,27]. Temperature stress could be categorized in to different type one is heat stress and cold stress. Although both kind of stress equally hampers plant physiology and biochemical mechanism. Heat stress sometime stimulates early flowering hence causing the start of reproduction process before time similarly the cold stress causes delay of flowering delaying the formation of seeds. Temperature stress sometime asynchronizes the maturation of male and female part [33].

In barley root tip, no activity of GPX on temperature stress [76]. When 2-year-old apple (*Malus domestica* Borkh) exposed at 40°C for 0-8hr, GPX activity was highest until 4 hr after that starting decreasing [79].

Heavy metal stress – In this present era, with growing industrialization and technology has led to release of toxic heavy metal elements such as Iron, Arsenate, Cadmium, Chromium, Lead, Copper, Mercury and Aluminum in environment has created global threat for all human beings [8]. This metal when found in trace amount are useful to life [8,28] but when accumulated in excessive amounts in plant causes reducing fertility and other damaging impact on it [29,30,31]. This chemicals through industrial effluent gets in contact with the soil, accumulates in it and then toxicating the soil. Increase in quantity of heavy metal in cellular level causes damage in mechanisms of plant, one of the most common ones is overproduction of ROS (reactive oxygen species) causing oxidative stress. Other harmful effects such as inactivation of biomolecules by displacing essential metal ions or obstructing functional groups [8,33]

In barley root tips – Cd-induced inhibits growth of root with increased activity of GPX, slight increase is observed when treated with Pb, Ni, and Zn, strong increase when treated with Hg and Cu and no activity when treated with Co[76].

3. DISCUSSION

Different type of abiotic stress and their detrimental effects on plants development and their productivity is the major concerning issue for researchers. When exposed to certain abiotic stress, plants response against those stresses are quite complex signalling pathways [8]. Plant has evolved various methods for sustaining under this constant physiological stress. The significant increase in the ROS (Reactive Oxygen Species) and RNS (Reactive Nitrogen Species) during the various abiotic stresses impose severe threat towards the survivability of these habitats. ROS and RNS can cause severe damage in cells by lipid Peroxidation, Oxidation of proteins and DNA, ultimately causing death of the cell (Parida et al., 2004). Antioxidant enzymes are one of the major defense mechanisms for plants to counter the detrimental effects of the reactive species generated in them (Dasgupta et al., 2012). Many scientific approaches had been discovered for proper genetic expression studies of such genes which helped them to determine and come up with all the mechanisms that have been helping plants from eons to protect themselves from such stresses. The fact that how antioxidant enzymes acting as a first line of defense and maintaining homeostasis of ROS in plant had proven a boon to us. Further researches and studies providing us with more new information and with their every achievement we are moving one step forward towards the insight of how we can protect more plants by inducing this mechanism in order to survive in the worsening situation of environment which we might face in future.

References

- [1.] Arora B.B., Sabhrawal A.K. (2013-2014) Digestion and Absorption. In: Ruchi Arora(ed), Ankur Sabhrawal(ed), Suman(ed) Modern's abc of Biology, Modern Publishers, Jalandhar City, pp 795-846
- [2.] Suzuki, N., Rivero, R. M., Shulaev, V., Blumwald, E., & Mittler, R. (2014). Abiotic and biotic stress combinations. *New Phytologist*, 203(1), 32-43

- [3.] Mittler, Ron. "Abiotic stress, the field environment and stress combination." *Trends in plant science* 11.1 (2006): 15-19.
- [4.] Choudhury, F. K., Rivero, R. M., Blumwald, E., & Mittler, R. (2017). Reactive oxygen species, abiotic stress and stress combination. *The Plant Journal*, 90(5), 856-867.
- [5.] Zhu, Jian-Kang. "Abiotic stress signaling and responses in plants." *Cell* 167.2 (2016): 313-324.
- [6.] Pereira, Andy. "Plant abiotic stress challenges from the changing environment." *Frontiers in plant science* 7 (2016): 1123.
- [7.] Li, Robert, Zhenquan Jia, and Michael A. Trush.
 "Defining ROS in biology and medicine." *Reactive oxygen* species (Apex, NC) 1.1 (2016): 9.
- [8.] Anwar, Ali, and Ju-Kon Kim. "Transgenic breeding approaches for improving abiotic stress tolerance: Recent progress and future perspectives." *International journal of molecular sciences* 21.8 (2020): 2695
- [9.] Cavanagh C, Morell M, Mackay I, Powell W. 2008. From mutations to MAGIC: resources for gene discovery, validation and delivery in crop plants. Current Opinion in Plant Biology 11: 215–221.
- [10.] Munns R, TesterM. 2008.Mechanisms of salinity tolerance. Annual Review of Plant Biology 59: 651–681.
- [11.] Chinnusamy V, Zhu JK. 2009. Epigenetic regulation of stress responses in plants. Current Opinion in Plant Biology 12: 133–139
- [12.] Mittler R, Blumwald E. 2010. Genetic engineering for modern agriculture: challenges and perspectives. Annual Review of Plant Biology 61: 443–462.
- [13.] Farooq, Muhammad, et al. "Drought stress in plants: an overview." *Plant responses to drought stress: From morphological to molecular features* (2012): 1-33.
- [14.] Lambers H, Chapin FS, Pons TL (2008) Plant physiological ecology, 2nd edn. Springer, New York
- [15.] Farooq M, Wahid A, Lee D-J (2009d) Exogenously applied polyamines increase drought tolerance of rice by improving leaf water status, photosynthesis and membrane properties. Acta Physiol Plant 31:937–945
- [16.] Mishra, V., & Cherkauer, K. A. (2010). Retrospective droughts in the crop growing season: Implications to corn and soybean yield in the Midwestern United States. Agricultural and Forest Meteorology, 150(7-8), 1030-1045.
- [17.] Farooq M, Bramley H, Palta JA, Siddique KHM (2011) Heat stress in wheat during reproductive and grain filling phases. Crit Rev Plant Sci 30:491–507

- [18.] Singh B, Usha K (2003) Salicylic acid induced physiological and biochemical changes in wheat seedlings under water stress. Plant Growth Reg 39:137–14
- [19.] Li YP, Ye W, Wang M, Yan XD (2009) Climate change and drought: a risk assessment of cropyield impacts. Climate Res 39:31–46
- [20.] Yamaguchi-Shinozaki K, Shinozaki K (2006) Transcriptional regulatory networks in cellular responses and tolerance to dehydration and cold stresses. Annu Rev Plant Biol 57:781–803
- [21.] Negrão, Sónia, S. M. Schmöckel, and M. J. A. O. B. Tester. "Evaluating physiological responses of plants to salinity stress." *Annals of botany* 119.1 (2017): 1-11.
- [22.] Qadir M, Quillerou E, Nangia V, et al. 2014. Economics of salt-induced land degradation and restoration. Natural Resources Forum 38: 282–295
- [23.] Anwar, A.; Liu, Y.; Dong, R.; Bai, L.; Yu, X.; Li, Y. The physiological and molecular mechanism of brassinosteroid in response to stress: A review. Biol. Res. 2018, 51, 46.
- [24.] Kumar, M.; Choi, J.; Kumari, N.; Pareek, A.; Kim, S. Molecular breeding in Brassica for salt tolerance: Importance of microsatellite (SSR) markers for molecular breeding in Brassica. Front. Plant Sci. 2015, 6, 688
- [25.] Gill, S.S.; Tuteja, N. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. Plant Physiol. Biochem. 2010, 48, 909–930
- [26.] Balazadeh, S.; Siddiqui, H.; Allu, A.D.; Matallana-Ramirez, L.P.; Caldana, C.; Mehrnia, M.; Zanor, M.-I.; Köhler, B.; Mueller-Roeber, B. A gene regulatory network controlled by the NAC transcription factor ANAC092/AtNAC2/ORE1 during salt-promoted senescence. Plant J. 2010, 62, 250–264.
- [27.] Liu, Q.; Yan, S.; Yang, T.; Zhang, S.; Chen, Y.-Q.; Liu, B. Small RNAs in regulating temperature stress response in plants. J. Integr. Plant Biol. 2017, 59, 774–791
- [28.] Sharma, P.; Anil, K.; Bhardwaj, R. Plant steroidal hormone epibrassinolide regulate—Heavy metal stress tolerance in Oryza sativa L. by modulating antioxidant defense expression. Environ. Exp. Bot. 2016, 122, 1–9
- [29.] Ali, B.; Hasan, S.A.; Hayat, S.; Hayat, Q.; Yadav, S.; Fariduddin, Q.; Ahmad, A. A role for brassinosteroids in the amelioration of aluminium stress through antioxidant system in mung bean (Vigna radiata L. Wilczek). Environ. Exp. Bot. 2008, 62, 153–159
- [30.] Jalmi, S.K.; Bhagat, P.K.; Verma, D.; Noryang, S.; Tayyeba, S.; Singh, K.; Sharma, D.; Sinha, A.K. Traversing the links between heavy metal stress and plant signaling. Front. Plant Sci. 2018, 9, 12.

- [31.] Lešková, A.; Giehl, R.F.H.; Hartmann, A.; Fargašová, A.; von Wirén, N. Heavy metals induce iron deficiency responses at different hierarchic and regulatory levels. Plant Physiol. 2017, 174, 1648–1668.
- [32.] Zinn, Kelly E., Meral Tunc-Ozdemir, and Jeffrey F. Harper. "Temperature stress and plant sexual reproduction: uncovering the weakest links." *Journal of experimental botany* 61.7 (2010): 1959-1968.
- [33.] Gill, S.S.; Tuteja, N. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. Plant Physiol. Biochem. 2010, 48, 909–930.
- [34.] Ray PD, Huang BW, Tsuji Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. Cell Signal. 2012 May;24(5):981-90. doi: 10.1016/j.cellsig.2012.01.008. Epub 2012 Jan 20. PMID: 22286106; PMCID: PMC3454471.
- [35.] Trachootham, D., Alexandre, J., & Huang, P. (2009). Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach?. *Nature reviews Drug discovery*, 8(7), 579-591.; 8:579–591. [PubMed: 19478820
- [36.] Andersen, J. K. (2004). Oxidative stress in neurodegeneration: cause or consequence?. *Nature medicine*, *10*(Suppl 7), S18-S25.
- [37.] Shukla, V., Mishra, S. K., & Pant, H. C. (2011). Adv Pharmacol Sci, 572634.
- [38.] Paravicini, T. M., & Touyz, R. M. (2006). Redox signaling in hypertension. *Cardiovascular research*, 71(2), 247-258.
- [39.] Haigis, M. C., & Yankner, B. A. (2010). The aging stress response. *Molecular cell*, 40(2), 333-344.
- [40.] Rajput, Vishnu D., et al. "Recent developments in enzymatic antioxidant defence mechanism in plants with special reference to abiotic stress." *Biology* 10.4 (2021): 267.
- [41.] Kasote, Deepak M., et al. "Significance of antioxidant potential of plants and its relevance to therapeutic applications." *International journal of biological sciences* 11.8 (2015): 982.
- [42.] Herbette, S.; Labrouhe, D.T.d.; Drevet, J.R.; Roeckel-Drevet, P. Transgenic tomatoes showing higher glutathione peroxydase antioxidant activity are more resistant to an abiotic stress but more susceptible to biotic stresses. Plant Sci. 2011, 180, 548–55
- [43.] Rajput, V.D.; Minkina, T.; Yaning, C.; Sushkova, S.; Chapligin, V.A.; Mandzhieva, S. A review on salinity adaptation mechanism and characteristics of Populus

euphratica, a boon for arid ecosystems. Acta Ecol. Sin. 2016, 36, 497–503

- [44.] Miller, G.A.D.; Suzuki, N.; Ciftci-Yilmaz, S.; Mittler, R.O.N. Reactive oxygen species homeostasis and signalling during drought and salinity stresses. Plant Cell Environ. 2010, 33, 453–467.
- [45.] Bartels, D.; Sunkar, R. Drought and Salt Tolerance in Plants. Crit. Rev. Plant Sci. 2005, 24, 23–58.
- [46.] Moussa, Ziad, Z. M. Judeh, and Saleh A. Ahmed. "Nonenzymatic exogenous and endogenous antioxidants." *Free radical medicine and biology* 1 (2019): 11-22.
- [47.] Zhou, Y.; Hu, L.; Ye, S.; Jiang, L.; Liu, S. Genome-wide identification of glutathione peroxidase (GPX) gene family and their response to abiotic stress in cucumber. 3 Biotech 2018, 8, 159.
- [48.] Zhou, Y.; Hu, L.; Wu, H.; Jiang, L.; Liu, S. Genome-wide identification and transcriptional expression analysis of cucumber superoxide dismutase (SOD) family in response to various abiotic stresses. Int. J. Genom. 2017, 2017, 7243973
- [49.] . Rajput, V.D.; Chen, Y.; Ayup, M. Effects of high salinity on physiological and anatomical indices in the early stages of Populus euphratica growth. Russ. J. Plant Physiol. 2015, 62, 229–236
- [50.] Yoshida K, Kaothien P, Matsui T, Kawaoka A, Shinmyo A. Molecular biology and application of plant peroxidase genes. Appl Microbiol Biotechnol. 2003 Feb;60(6):665-70. doi: 10.1007/s00253-002-1157-7. Epub 2002 Dec 18. PMID: 12664144.
- [51.] Hiraga, Susumu, et al. "A large family of class III plant peroxidases." *Plant and Cell Physiology* 42.5 (2001): 462-468.
- [52.] Churin, Y., Schilling, S. and Borner, T.(1999) FEBS Lett. 459: 33-38
- [53.] Welinder, K.G. (1992) Curr. Opin. Stuct. Biol. 2:388-393
- [54.] Bela, Krisztina, et al. "Plant glutathione peroxidases: emerging role of the antioxidant enzymes in plant development and stress responses." *Journal of Plant Physiology* 176 (2015): 192-201.
- [55.] Toppo S, Flohe L, Ursini F, Vanin S, Maiorino M. Catalytic mechanisms and specificities of glutathione peroxidases: variations of a basic scheme. Biochim Biophys Acta 2009;1790:1486–500.
- [56.] Mullineaux PM, Karpinski S, Jiménez A, Cleary SP, Robinson C, Creissen GP. Identification of cDNAS encoding plastid-targeted glutathione peroxidase. Plant J 1998;13:375–9.

- [57.] Yang XD, Li WJ, Liu JY. Isolation and characterization of a novel PHGPx gene in Raphanus sativus. Biochim Biophys Acta 2005;1728:199–205
- [58.] Roxas VP, Smith RK Jr, Allen ER, Allen RD. Overexpression of glutathione Stransferase/glutathione peroxidase enhances the growth of transgenic tobacco seedlings during stress. Nat Biotechnol 1997;15:988–91
- [59.] Csiszár J, Szabó M, Erdei L, Márton L, Horváth F, Tari I. Auxin autotrophic tobacco callus tissues resist oxidative stress:the importance of glutathione S-transferase and glutathione peroxidase activities in auxin heterotrophic and autotrophic calli. J Plant Physiol 2004;161:691–9.
- [60.] Kilili KG, Atanassova N, Vardanyan A, Clatot N, Al-Sabarna K, Kanellopoulos PN, et al. Differential roles of Tau class glutathione S-transferases in oxidative stress. J Biol Chem 2004;279:24540–51
- [61.] Basantani M, Srivastava A. Plant glutathione transferases—a decade falls short. Can J Bot 2007;85:443–56.
- [62.] Dixon DP, Hawkins T, Hussey PJ, Edwards R. Enzyme activities and subcellular localization of members of the Arabidopsis glutathione transferase superfamily. J Exp Bot 2009;60:1207–18
- [63.] EdwardsR, Dixon DP. Selective binding of glutathione conjugates of fatty acid derivatives by plant glutathione transferases. J Biol Chem 2009;284:21249–56.
- [64.] Navrot N, Collin V, Gualberto J, Gelhaye E, Hirasawa M, Rey P, et al. Plant glutathione peroxidases are functional peroxiredoxins distributed in several subcellular compartments and regulated during biotic and abiotic stresses. Plant Physiol 2006;142:1364–79
- [65.] Koh CS, Didierjean C, Navrot N, Panjikar S, Mulliert G, Rouhier N, et al. Crystal structures of a poplar thioredoxin peroxidase that exhibits the structure of glutathione peroxidases: insights into redox-driven conformational changes. J Mol Biol 2007;370:512–29
- [66.] Herbette S, Lenne C, Leblanc N, Julien JL, Drevet JR, Roeckel-Drevet P. Two GPX-like proteins from Lycopersicon esculentum and Helianthus annuus are antioxidant enzymes with phospholipid hydroperoxide glutathione peroxidase and thioredoxin peroxidase activities. Eur J Biochem 2002;269:2414–20
- [67.] Rodriguez Milla MA, Maurer A, Rodriguez Huete A, Gustafson JP. Glutathione peroxidase genes in Arabidopsis are ubiquitous and regulated by abiotic stresses through diverse signaling pathways. Plant J 2003;36:602–15

- [68.] Pandey, Veda P., et al. "A comprehensive review on function and application of plant peroxidases." *Biochem Anal Biochem* 6.1 (2017): 308.
- [69.] Yuchen Miao, Dong Lv, Pengcheng Wang, Xue-Chen Wang, Jia Chen, Chen Miao, Chun-Peng Song, An Arabidopsis Glutathione Peroxidase Functions as Both a Redox Transducer and a Scavenger in Abscisic Acid and Drought Stress Responses, *The Plant Cell*, Volume 18, Issue 10, October 2006, Pages 2749–2766
- [70.] Mittova, V., Theodoulou, F. L., Kiddle, G., Gómez, L., Volokita, M., Tal, M., et al. (2003). Coordinate induction of glutathione biosynthesis and glutathione-metabolizing enzymes is correlated with salt tolerance in tomato. *FEBS Lett.* 554, 417–421. doi: 10.1016/S0014-5793(03)01214-6
- [71.] Zhang, Lipeng, et al. "Overexpression of the glutathione peroxidase 5 (RcGPX5) gene from rhodiola crenulata increases drought tolerance in Salvia miltiorrhiza." Frontiers in Plant Science 9 (2019): 1950.
- [72.] Zhu, J.K. (2002). Salt and drought stress signal transduction in plants. Annu. Rev. Plant Biol. 53, 247–273
- [73.] Park, S.Y., Fung, P., Nishimura, N., Jensen, D.R., Fujii, H., Zhao, Y., Lumba, S., Santiago, J., Rodrigues, A., Chow, T.F., et al. (2009). Abscisic acid inhibits type 2C protein phosphatases via the PYR/PYL family of START proteins. Science 324, 1068–1071
- [74.] Ma, Y., Szostkiewicz, I., Korte, A., Moes, D., Yang, Y., Christmann, A., and Grill, E. (2009). Regulators of PP2C phosphatase activity function as abscisic acid sensors. Science 324, 1064–1068
- [75.] Sharma, Pallavi, and Rama Shanker Dubey. "Drought induces oxidative stress and enhances the activities of antioxidant enzymes in growing rice seedlings." *Plant* growth regulation 46 (2005): 209-221.
- [76.] Halušková, L'ubica, et al. "Effect of abiotic stresses on glutathione peroxidase and glutathione S-transferase activity in barley root tips." *Plant Physiology and Biochemistry* 47.11-12 (2009): 1069-1074.
- [77.] Sadak, Mervat, Ebtihal Abd Elhamid, and Marwa Mahmoud. "Glutathione induced antioxidant protection against salinity stress in chickpea (Cicer arietinum L.) plant." *Egyptian Journal of Botany* 57.2 (2017): 293-302.
- [78.] Paiva, Ana Luiza S., et al. "Mitochondrial glutathione peroxidase (OsGPX3) has a crucial role in rice protection against salt stress." *Environmental and Experimental Botany* 158 (2019): 12-21.
- [79.] Ma, Yu-Hua, et al. "Effects of high temperature on activities and gene expression of enzymes involved in

ascorbate–glutathione cycle in apple leaves." *Plant Science* 175.6 (2008): 761-766.

- [80.] Dasgupta, N., Nandy, P., Tiwari, C., & Das, S. (2010). Salinity-imposed changes of some isozymes and total leaf protein expression in five mangroves from two different habitats. *Journal of Plant Interactions*, 5(3), 211-221.
- [81.] Dasgupta, N., Nandy, P., & Das, S. (2011). Photosynthesis and antioxidative enzyme activities in five Indian mangroves with respect to their adaptability. *Acta physiologiae plantarum*, 33, 803-810.
- [82.] Parida A.K., Das A.B., Mohanty P. 2004b. Investigations on the antioxidative defense responses to NaCl stress in a mangrove, *Bruguiera parviflora*: Differential regulations of isoforms of some antioxidative enzymes. Plant Growth Regulat. 42(3):213–226
- [83.] Dasgupta, N., Nandy, P., Sengupta, C., & Das, S. (2012). Protein and enzymes regulations towards salt tolerance of some Indian mangroves in relation to adaptation. *Trees*, 26, 377-391.

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